文章编号:1000-4025(2007)12-2491-05

外源 HeO2 对小麦幼苗耐盐性的调节作用

张 波1,2,张怀刚1*

(1 中国科学院 西北高原生物研究所,西宁 810001;2 中国科学院 研究生院,北京 100049)

摘 要:以 高原 448 '小麦品种为材料,分别测定了 4 个处理 (Hoagland 营养液、Hoagland 营养液 + 150 mmol/L NaCl、Hoagland 营养液 + 150 mmol/L NaCl + 10 μ mol/L H₂O₂ 和 Hoagland 营养液 + 10 μ mol/L H₂O₂) 的小麦幼苗在第 2、4、6、8 天叶片叶绿素、丙二醛、可溶性糖和还原性谷胱甘肽含量. 结果显示:外源 H₂O₂ 提高了 NaCl 胁迫下 4 个时段小麦幼苗的叶绿素含量 (8. 27 %、32. 57 %、10. 19 %、4. 86 %) 及还原性谷胱甘肽含量 (3. 09 %、23. 97 %、5. 85 %、2. 11 %),显著提高了可溶性糖含量 (14. 58 %、8. 43 %、16. 68 %、5. 8 %,P < 0.05),而显著降低了其丙二醛含量 (17. 53 %、14. 04 %、4. 75 %、8. 47 %,P < 0.05).外源 H₂O₂ (10 μ mol/L) 使 NaCl 胁迫下叶绿素含量和还原性谷胱甘肽含量峰值提前,同时推迟了丙二醛峰值出现的时间。研究表明,外源 H₂O₂ 通过提高叶片叶绿素、可溶性糖和还原性谷胱甘肽含量以有效地增强小麦幼苗的耐盐性.

关键词:过氧化氢;小麦幼苗;耐盐性;调节作用中图分类号:Q945.78 文献标识码:A

Regulation of Exogenous Hydrogen Peroxide on Wheat Seedling Salinity Tolerance

ZHANG Bo^{1,2}, ZHANG Huai-gang^{1*}

(1 Northwest Institute of Plateau Biology, Chinese Academy of Sciences, Xining 810001, China; 2 Graduate University, Chinese Academy of Sciences, Beijing 100049, China)

Abstract: The chlorophyll content ,MDA content ,soluble sugar content and GSH content were determined at the 2^{nd} , 4^{th} , 6^{th} and 8^{th} day ,respectively ,after treatments of Hoagland solution , Hoagland solution + 150 mmol/L NaCl , Hoagland solution + 150 mmol/L NaCl , Hoagland solution + 150 mmol/L H_2O_2 and Hoagland solution + 10 μ mol/L H_2O_2 . The addition of exogenous H_2O_2 (10 μ mol/L) increased the wheat seedling chlorophyll content (by 8. 27 % ,32.57 % ,10.19 % and 4.86 %) and GSH contents (by 3.09 % ,23.97 % ,5.85 % and 2.11 %) ,dramatically increased soluble sugar content (by 14.58 % ,8.43 % ,16.68 % and 5.8 % , P < 0.05) and dramatically decreased MDA content (by 17.53 % ,14.04 % ,4.75 % and 8.47 % , P < 0.05) compared to the salt stressed (Hoagland solution + NaCl) group on the 2^{nd} , 4^{th} , 6^{th} and 8^{th} day ,respectively ,after treatments. Exogenous H_2O_2 (10 μ mol/L) accelerated the maximum accumulation chlorophyll and GSH contents in NaCl stressed condition while decelerated the maximum accumulation content of MDA. Exogenous H_2O_2 enhanced wheat seedling salinity tolerance.

Key words: H₂O₂; wheat seedling; salinity tolerance; regulation

Soil salinity is one of the major abiotic stresses affecting plant growth and productivity globally.

^{*}收稿日期:2007-04-30;修改稿收到日期:2007-10-25

基金项目:中国科学院知识创新工程重要方向性项目(KSCX2-SW-304);中科院中组部西部之光联合学者项目资助

作者简介:张 波(1980-),男(汉族),在读博士研究生,主要从事植物抗逆生理生化及分子生物学研究.

^{*}通讯作者:张怀刚,博士,研究员,博士生导师,主要从事小麦遗传育种及生理生化研究. E-mail:bzhang6668 @sina.com

There are 380 million hectares of salinity soil in the world. One of third irrigable fields is affected by high salinity. In plants, hydrogen peroxide (H₂O₂) is thought to be increased under various abiotic stresses and to enhance gene expression of active oxygen scavenging (AOS) enzymes. In maize, H₂O₂ production was reported to increase in response to chilling stress, and exogenously applied H₂O₂ increased chilling tolerance^[1]. Increased H₂O₂ production was also reported to occur gradually in response to salt stress in rice plants^[2]. Pretreating rice seedlings with low level of H2O2 increased their salt tolerance^[3]. Wheat as a crop of great importance in the world, is one of the saltsensitive crops. Accumulation of salts in irrigated soil is one of the primary factors limiting yield in wheat production. Research on salt stress tolerance in wheat is therefore of great importance for future practical advances. However, little research has been devoted to the role of exogenous H2O2 on salt tolerance of wheat plants. In the present study, we demonstrated that exogenously applied H₂O₂ caused protection against salt in young wheat seedlings.

1 Materials and methods

1.1 Plant materials and treatments

Seeds used in the experiment were from spring wheat (Triticum aestivum L.) variety "Plateau 448". The variety was developed by Northwest Institute of Plateau Biology, the Chinese Academy of Sciences (CAS) and released in 1999. It has become one of the major cultivars in the Qinghai-Tibet Plateau and Northwest Spring Wheat Area. The seeds were surface sterilized with 70 % alcohol and thoroughly rinsed by tap water and then transferred to two sheets of sterilized moist filter paper. Seeds were placed in plastic trays for germination at 28 for 48 h in the dark. Uniform germinated seeds were selected and sown into the culture dishes, and grown hydroponically in a growth chamber. The temperature of the growth chamber was maintained at (27 ± 2) and light intensity was 400 µmol• m⁻²• s⁻¹. The humidity of the growth chamber was between 60 % ~ 70 %. The Hoagland nutrition was aerated continuously and replaced every two days. When the third leaf sprouted ,treatments were made as follows:(1) Hoagland solution;(2) Hoagland solution + 150 mmol/L NaCl; (3) Hoagland solution + 150 mmol/L NaCl + 10 μ mol/L H₂O₂;(4) Hoagland solution + 10 μ mol/L H₂O₂. Leaves at the same leaf order were harvested at 9 O'clock on the 2^{nd} , 4^{th} , 6^{th} and 8^{th} day after treatment , respectively and then stored at - 70 for determination of chlorophyll content ,MDA content ,soluble sugar content and GSH content. All the treatments were performed in triplicates.

1.2 Methods

Chlorophyll content was determined as described in Yang 's method^[4]. MDA and soluble sugar content were determined following Ellman 's method^[5]. Reduced glutathione (GSH) content was determined following Ellman 's method^[6]. All the determinations were accomplished by 751-UV spectrophotometer and performed in triplicates and the average values were presented. Excel and SPSS15.0 were used for data statistic analysis.

2 Results

2.1 Effects of exogenous H_2O_2 on chlorophyll content in wheat seedling

As we all know, photosynthesis is one of the key metabolic pathways that are responsible for growth and development of plants^[7] and chlorophyll is the most important photosynthetic pigment in plants. Therefore chlorophyll content could be used as an index of salt tolerance of plants. As shown in Fig. 1, after wheat seedlings were grown in 150 mmol/L NaCl, the leaf chlorophyll content was decreased by 13.58%, 12.82%, 3.27% and 1.82 % on the 2nd, 4th, 6th, 8th day, respectively, compared to the control (Hoagland solution treated only). While the chlorophyll content of those groups with Hoagland + NaCl + H2O2 treated was much higher than that of the groups with salt stressed (150 mol/L NaCl). The addition of $H_2\,O_2$ (10 µmol/L) to salt stressed wheat seedlings increased their chlorophyll contents by 8.27 %, 32. 57 %, 10.19 % and 4.86 % on the 2nd, 4th, 6th and 8th day respectively. Especially on the 4th, 6th, and 8th

day after treatments, the chlorophyll content of Hoagland + NaCl + H_2O_2 treated group was even higher than that of the Hoagland treated group. On the 2^{nd} and 8^{th} day after treatment of Hoagland + H_2O_2 , the chlorophyll content in wheat leaves was a little higher than control. Furthermore, the maximum chlorophyll content appeared on the 6^{th} day in Hoagland + NaCl treated group while on the 4^{th} day in Hoagland + NaCl + H_2O_2 treated group.

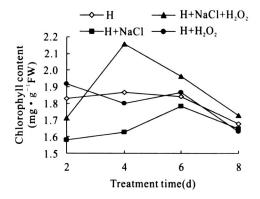


Fig. 1 Chlorophyll content in wheat seedling with different treatments and time

H represents Hoagland solution; Results are presented as means and standard errors from three replications.

The same as below

2.2 Effects of exogenous H_2O_2 on lipid peroxidation in wheat seedling

Lipid peroxidation of wheat seedlings after treatments, measured as MDA content, is shown in Fig. 2. After salt stressed treatment (150 mol/L NaCl), the MDA content was increased by 13.89 %, 27.63 %, 29.35 % and 19.67 % on the 2^{nd} , 4^{th} , 6^{th} and 8^{th} day, respectively, compared with the control. While the MDA content was decreased by 17.53 %, 14.04 %, 4.75%, 8.47% in the Hoagland + NaCl + H₂O₂ on the 2^{nd} , 4^{th} , 6^{th} and 8^{th} day after treatments (P <0.05) ,respectively ,compared with the salt stressed group. In addition, on the 2nd, 4th, 8th day after treatments, the MDA content of the Hoagland + H₂O₂ treated group was decreased a little compared to the control. MDA content was increased before 4th and decreased during the 4th ~ 8th day after treatment of Hoagland + NaCl. While the MDA content was increased until the 6^{th} day and then decreased. Exogenously added H₂O₂ not only decreased MDA content but decelerated the appearance of maximum MDA content in NaCl stressed condition.

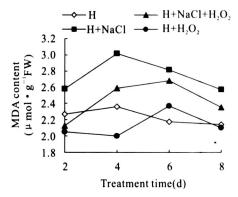


Fig. 2 MDA content in wheat seedling with different treatments and time

2.3 Effects of exogenous H_2O_2 on soluble sugar content in wheat seedling

Soluble sugar content is shown in Fig. 3. Both salt stressed treatment (Hoagland + NaCl) and Hoagland + NaCl + H₂O₂ treatment increased the wheat seedling soluble sugar content to different degrees on the 2nd, 4th ,6th and 8th day after treatments. The stressed treatment (Hoagland + NaCl) increased soluble sugar content by 22.38 %,11.66 %,8.41 %,7.07 % on the 2nd, 4th ,6th and 8th day after treatments, respectively, compared with the control. While the soluble sugar contents of Hoagland + NaCl + H₂O₂ treated group were increased by 36.96 %, 20.09 %, 25.09 %, 12.87 % on the 2nd ,4th ,6th and 8th day ,respectively ,compared with the control. Hoagland + NaCl + H₂O₂ treatment further increased the soluble sugar by 14.58 %, 8.43 %, 16.68 % and 5.8 %, respectively, compared to the salt stressed group (P < 0.05). Soluble sugar contents were increased from 2nd to 4th day and then decreased during the next 4 days in the all treatments except for the Hoagland + H₂O₂ treated group.

2.4 Effects of exogenous H₂O₂ to GSH content of wheat seedling

GSH content is displayed in Fig. 4. Salt stress treatment (Hoagland + NaCl) decreased the GSH content by 8. 68 % ,20. 27 % ,10. 06 % and 8. 62 % on the 2^{nd} ,4th ,6th and 8th day ,respectively ,compared with the control. The addition of H_2O_2 (10 μ mol/L) to Hoagland + NaCl increased the GSH content by 3. 09 % , 23. 97 % ,5. 85 % and 2. 11 % , respectively ,compared with the salt stressed group at the 2^{nd} ,4th ,6th ,8th day after treatments. GSH content was increased more dra-

matically in Hoagland + NaCl + $H_2 \, O_2$ treated group than in Hoagland + NaCl treated group from 2^{th} to 4^{th} day after treatment. The appearance of maximum GSH content was also accelerated by exogenously added $H_2 \, O_2$ in NaCl stressed condition.

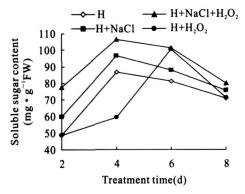


Fig. 3 Soluble sugar content in wheat seedling with different treatments and time

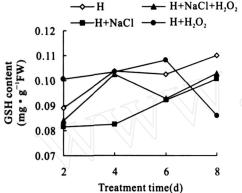


Fig. 4 GSH content in wheat seedling with different treatments and time

3 Discussion

Compounds that are able to reduce damaging effects of various stresses should be of great importance from both the theoretical and application points of view. The ROS hydrogen peroxide (H_2O_2) has generally been viewed as a toxic cellular metabolite. However, it is clear that it may also function as a signal molecule in both plant and animal cells^[8]. The generation of H_2O_2 is increased in response to a wide variety of abiotic and biotic stresses, and some authors have suggested that H_2O_2 plays a dual role in plants:at low concentrations, it acts as a messenger molecule involved in acclimatory signaling, triggering tolerance against various abiotic stresses, and at high concentrations it orchestrates programmed cell death^[1,9]. Thus, it appears likely that H_2O_2 accumulation in specific tissues,

and in the appropriate quantities, may benefit plants by mediating plant acclimation and cross-tolerance to both biotic and abiotic stresses^[9].

In the present study we showed that exogenous $H_2\,O_2$ caused protection against salt stress in wheat young seedlings (Fig. 1 ~ 4) . Chlorophyll content could be used as an index of salt tolerance of plants. Decreasing of wheat seedling chlorophyll content caused by salt stress (150 mmol/L NaCl) was alleviated by the $H_2\,O_2$ (10 μ mol/L) to different degrees. Hoagland + $H_2\,O_2$ (10 μ mol/L) treatment also increased the wheat seedling chlorophyll content of Hoagland + $H_2\,O_2$ group to the level of being beyond the control on the 4^{th} , 6^{th} and 8^{th} day after treatment. At the same time, the appearance of maximum chlorophyll content was accelerated by exogenously added $H_2\,O_2$ (Fig. 1). That indicated the exogenous $H_2\,O_2$ could play some protective role in salt stress condition.

Salt stress is known to result in extensive lipid peroxidation ,which has often been used as indicator of salt-induced oxidative membrane damages^[8]. MDA is an end product of lipid peroxidation. In the present study (Fig. 2) ,we found that MDA content was decreased by the exogenously added H_2O_2 (10 μ mol/L) compared with salt stressed group. We can see clearly from the data that the protection caused by H_2O_2 against salt stress occurred mainly in the early stage after treatment of H_2O_2 (10 μ mol/L). It also showed that H_2O_2 had little effect on unstressed wheat seedlings. Exogenously added hydrogen peroxide not only decreased MDA content but decelerated the appearance of maximum MDA content in the NaCl stressed condition.

Water deficiency caused by salt stress is a common phenomenon in plants. Water deficiency often leads to metabolic disorders. In order to keep a relatively higher osmotic potential ($_{\rm s}$), plants often positively synthesize and accumulate some compatible solutes which could increase the osmotic potential of plant cells. Soluble sugar is one of compatible solutes. In order to protect plant cells from losing water excessively, soluble sugar content was increased under salt stressed condition, while the addition of H_2O_2 (10 μ mol/L) further in-

creased the accumulation of soluble sugar in wheat seedlings (Fig. 3) ,hence protected wheat seedling cells by keeping a relatively higher osmotic potential ($_{\rm s}$) and improving their salt tolerance. It was showed that at the $2^{\rm nd}$ and $6^{\rm th}$ day after treatment of Hoaglan + $H_2\,O_2$ (10 μ mol/L), the soluble sugar content was also higher than control (Fig. 3). So we could speculate it is the "Pie-up Effect" of salt stress and $H_2\,O_2$.

GSH is an important metabolite as it is involved in the protection of plants from oxidative stress buildup due to various environmental stresses $^{[10^{-12}]}$. GSH is one of the non-enzymatic antioxidants like ascorbate (ASA) ,tocopherols and carotenoids for scavenging reactive oxygen species (ROS) produced in the stressed condition. One of the damages caused by the salt stress is the oxidative stress. Our work showed that salt stress decreased the GSH content while the addition of H_2O_2 (10 μ mol/L) increased the GSH content compared to the salt stressed treatment, hence increased the salt tolerance of wheat seedlings. The appearance of maximum GSH content was also ac-

celerated by exogenously added H_2O_2 in NaCl stressed condition. In addition, H_2O_2 (10 μ mol/L) also increased the GSH content of unstressed group, but its influences on unstressed group was much less than that of salt stressed group.

We found that H_2O_2 as a newly discovered signal molecule was much more effective in salt stressed condition than in unstressed condition. It could be hypothesized that H₂O₂ may induce the expression of series of related defense genes immediately when salt stress occurs. It is reported that endogenous H₂O₂ accumulation occurs at the early stage ,providing evidence that H₂O₂ might be an inducer of antioxidation mechanism^[1]. In our experiment, exogenous H₂O₂ was added and the 2nd, 4th, 6th and 8th day after treatments were selected for the determination of physiological indexes. We also hypothesize that H₂O₂ regulate the physiology and biochemical metabolism mainly during the few hours after treatment. Therefore, it is worth understanding the effects of H₂O₂ on salt tolerance during the few hours after treatment in the future.

References:

- [1] PRASAD T K, ANDERSON M D, MARTIN B A, STEWART C R. Evidence for chilling-induced oxidative stress in maize seedlings and a regulatory role for hydrogen peroxide[J]. *Plant Cell*, 1994, 6:65 74.
- [2] FADZILLA N M, FINCH R P, BURDON R H. Salinity, oxidative stress and antioxidant responses in shoot cultures of rice [J]. J. Exp. Bot., 1997, 48:325 331.
- [3] UCHIDA A, ANDRE T, JAGENDORF, TAKASHI H, TERUHIRO T. Effects of hydrogen peroxide and nitric oxide on both salt and heat stress tolerance in rice[J]. Plant Sci., 2002, 163:515 523.
- [4] 杨善元. 叶绿素含量测定[A]. 见:汤章城. 现代植物生理学实验指南[M]. 北京:科学出版社, 1999:95-96.
- [5] 张雯. 植物组织 MDA 含量测定[A]. 见:张志良. 植物生理学实验指导[M]. 北京:高等教育出版社,2003:274 277.
- [6] ELLMAN GL. Tissue sulfhydryl groups[J]. Arch Bio. Chem. Biophys., 1959, 82:70 77.
- [7] WISE R R. Chilling-enhanced photooxidation: the production [J]. Photosyn. Res., 1995, 45:79 97.
- $[8] \quad FIN\,KEL\ T.\ Redox-dependent\ signal\ transduction [J]\ .\ FEBS\ Lett\ , 2000\ , 476:52-54.$
- [9] VAN BREUSEGEM F, VRANOVA 'E, DATJ F, INZE 'D. The role of active oxygen species in plant signal transduction[J]. *Plant Sci.*, 2001, 161:405 414.
- [10] BOWLER C, FIUHR R. The role of calcium and activated oxygens as signals for controlling cross-tolerance[J]. Trends Plant Sci., 2000, 5:241 246.
- [11] HERNA 'NDEZJ A ,ALMANSA M S. Short-term effects of salt stress on antioxidant systems and leaf water relations of pea leaves[J].

 Physiol. Plant ,2002 ,115:251 257.
- [12] MAY MJ, VERNOUX T, LEAVER C, VAN M M, INZE D. Glutathione homeostasis in plants: implications for environmental sensing and plant development [J]. J. Exp. Bot., 1998, 49:649 667.