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Low-level nitrogen deposition significantly inhibits methane uptake from an alpine meadow soil on the Qinghai–Tibetan Plateau



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ABSTRACT

It is crucial to understand the effects of enhanced nitrogen (N) deposition on soil methane (CH₄) uptake to develop a better comprehension of carbon (C) dynamics in terrestrial ecosystems. A two-year field study was conducted to assess the effects of various forms of N (NH_4^+ and NO_3^-) and associated N deposition rates (0, 10, 20 and 40 kg N ha⁻¹ yr⁻¹) on alpine meadow soil CH₄ fluxes on the Qinghai–Tibetan Plateau, China. Soil CH₄ fluxes, soil temperature, and soil moisture were monitored weekly using the static chamber technique and gas chromatography. Soil inorganic N pools, soil pH and aboveground biomass were measured monthly to examine the key controlling factors of soil CH₄ flux. Our results showed that N addition significantly promoted plant growth and changed soil water-filled pore space (WFPS), but did not alter soil inorganic N storages over the short term. Low rates of N addition significantly decreased the seasonal amount of CH₄ uptake by 8.6% compared with the control. Soil CH₄ fluxes were mainly determined by soil WFPS, followed by inorganic N availability. N addition increased the contribution of soil WFPS, pH and soil NO₃⁻ storage. The observed reduction in CH₄ uptake caused by N addition may be largely due to a decrease in physical diffusion, as the biochemical inhibition effects on methanotrophic bacteria are minor. These results suggest that soil inorganic N is a regulatory factor of soil CH₄ uptake, and its promotion or inhibition to soil CH₄ uptake depends on the N status in terrestrial ecosystems.

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1. Introduction

Methane (CH₄) is a powerful greenhouse gas and contributes to approximately 17% of global warming, with a relative global warming potential 23 times that of CO₂ on a molar basis (Parry, 2007). Over the past 200 years, human activities such as fossil fuel exploration, rice production, large-scale animal husbandry of ruminants, biomass burning and landfill gas emission have resulted in an increase of atmospheric CH₄ concentration by 148%, which is currently increasing by about 0.9% yr⁻¹ (Parry, 2007). Meanwhile, atmospheric CH₄ reacts with hydroxyl radicals (OH) in the troposphere and transfers to the stratosphere, accounting for 84% and 7% of the global CH₄ sink, respectively (Dutaur and Verchot, 2007). Also, uptake of CH₄ by aerobic soils removes a significant amount from the atmosphere (10–44 Tg yr⁻¹), and accounts for up to 10% of the global CH₄ sink (Lowe, 2006). However, there is evidence that increases of atmospheric nitrogen (N) deposition

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0016-7061/\$ – see front matter © 2013 Elsevier B.V. All rights reserved. http://dx.doi.org/10.1016/j.geoderma.2013.08.006 can significantly decrease CH₄ uptake by soils (Liu and Greaver, 2009), which may also contribute to increasing atmospheric CH₄ concentrations.

N addition may increase (Saari et al., 2004; Veldkamp et al., 2001), decrease (Domingues et al., 2007: Liebig et al., 2008: Mosier et al., 1998) or have no effects on (Phillips and Podrebarac, 2009; Sawamoto et al., 2010; Van den Pol-van Dasselaar et al., 1999) CH₄ uptake from grassland ecosystems, appearing to depend on the form and the rate of N addition, as well as on soil properties (Rigler and Zechmeister-Boltenstern, 1999). Serious debates have focused on the role of NH₄⁺ and NO₃⁻ on soil CH₄ consumption. Many studies have demonstrated that elevated soil NH⁺₄ may significantly reduce CH₄ oxidation rates (Bodelier and Laanbroek, 2004). However, several studies reported that NO_3^- , rather than NH_4^+ , had the greatest inhibitory effect on CH_4 oxidation in forest soils (Reay and Nedwell, 2004; Wang and Ineson, 2003; Xu and Inubushi, 2004; Xu and Inubushi, 2007). Unfortunately, most studies on the response of soil CH₄ uptake to the forms and rates of N addition have been conducted using soil core incubation in a laboratory, which differs from N addition experiments done in the field. Also, all of these studies were focused on temperate forest ecosystems (Reay and Nedwell, 2004; Wang and Ineson, 2003; Xu and Inubushi, 2004; Xu and Inubushi, 2007), with no data available



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from alpine meadow ecosystems. Overall, the patterns and intrinsic mechanisms of how NO_3^- and NH_4^+ affect CH_4 uptake in grassland ecosystems remain unclear.

As the largest grassland unit on the Eurasian continent, the Qinghai-Tibetan Plateau is averagely situated at 4000 m above sea level, and covers an area of approximately 2.5 million km² (Yang et al., 2008). Soil carbon (C) storage for 1 m in depth is estimated to be 4.68 Pg C, accounting for about 1/10 of total soil C storage in China (Yang et al., 2008). Due to an increase in regional economic development as well as long-range transport of atmospheric reactive N, atmospheric N deposition is very obvious in the eastern Qinghai-Tibetan Plateau, ranging from 4 to 13.8 kg N ha⁻¹ yr⁻¹ (Liu et al., 2013; Lü and Tian, 2007). Chronic N deposition can significantly change the soil CH₄ uptake over the short term (C. Jiang et al., 2010), and can further offset C sequestration in the entire alpine meadow ecosystem (Fang et al., 2012). To clarify CH₄ uptake in the alpine meadow system within the context of increased N deposition, it is necessary to explore the responses of soil CH₄ uptake by the different forms and the rates of N addition.

We hypothesized that NO_3^- and NH_4^+ played different roles to soil CH₄ uptake. This hypothesis was tested by conducting a two-year N addition experiment in an alpine meadow on the Qinghai–Tibetan Plateau. The main purposes of this paper are: (1) to examine the effects of the forms and rates of N addition on soil variables (soil temperature, water-filled pore space (WFPS), inorganic N pools and pH) and above-ground biomass; (2) to quantify the effects of the forms and rates of N addition on CH₄ uptake in the alpine meadow soil; and (3) to investigate which factors are responsible for changes in CH₄ uptake from the alpine meadow soil induced by N addition.

2. Materials and methods

2.1. Study site

The experiment was conducted at the Haibei alpine meadow ecosystem research station, Chinese Academy of Sciences ($37^{\circ}37'$ N, $101^{\circ}19'$ E), located in the eastern of Qinghai–Tibetan Plateau. The site is subjected to a plateau monsoon climate. Weather data collected between 2008 and 2009 at the meteorological station showed a mean annual air temperature of -0.4 °C, with an overall minimum and maximum of -23.2 °C (January) and 14.5 °C (July), respectively. The annual average precipitation was approximately 383.3 mm, of which 85.6% occurred in May and September (Fig. 1). The vegetation type is a typical Kobresia humilis meadow. Dominant species are Kobresia humilis, Saussurea superba, Potentilla saundersiana, Leontopodium nanum, Lancea tibetica, Festuca ovina, Festuca rubra, Stipa aliena, Elymus nutans, Helictotrichon tibetica, Koeleria cristata and Poa crymophila (Cao et al.,

30 40 Air temperature Precipitaiton Average precipitation (mm) Average temperature (°C) 20 30 10 20 0 -10 10 -20 0 -30 08-12 09-12 07-12 08-4 08-8 09-4 09-8 Date

Fig. 1. The annual patterns of the daily average temperature and precipitation of the study site in 2008 and 2009.

2008). The soils developed in the Kobresia meadow are Mat-Gryic Cambisol (Cao et al., 2008).

2.2. Experimental design

The N addition experiment is a split plot design with N levels defining the main plots and N forms as subplots. Three N fertilizers (NH₄Cl, (NH₄)₂SO₄ and KNO₃) were applied as three levels (10, 20 and 40) of kg N ha⁻¹ yr⁻¹. A control (0 kg N ha⁻¹ yr⁻¹) was set at each plot and each N treatment had three replicates. Each plot had an area of 9 m² (3 m × 3 m) and a 2 m isolation band was set between plots. The N addition experiment started on May 1, 2007. N fertilizer solutions were sprayed on the plots once a month, totally 12 equal applications over the year. Control plots received equivalent applications of water only. All plots are in the winter pastures, not grazed in the growing season, and grazed in non-growing season.

2.3. CH₄ flux measurement

Over the two growing seasons (from May to September) of 2008 and 2009, the soil CH₄ fluxes were determined using the static chamber and gas chromatography method (Wang and Wang, 2003). At each plot, a stainless square box (length \times width \times height = 50 cm \times 50 cm \times 50 cm), made up of one collar and one chamber, was installed. Gas samples were collected weekly between 9:00 AM and 11:00 AM (China Standard Time, CST) by fitting the chambers to the collars and removing headspace samples at ten-minute intervals over a 30-min period. Within 24 h following gas collection, the CH₄ concentrations were analyzed on a gas chromatograph (HP Series 4890D, Hewlett Packard, USA), equipped with a flame ionization detector (FID). The CH₄ fluxes were calculated, using chamber air temperature, barometric air pressure and the slope of the temporal change in CH₄ concentration within the chamber headspace (Wang and Wang, 2003). The seasonal amount of CH₄ uptake can be calculated using the following equation (J. Jiang et al., 2010):

$$E_{\text{CH4}} = \frac{\sum_{i=1}^{n} 0.5 \times (F_i + F_{i+1}) \times (t_{i+1} - t_i) \times 24}{1000}$$

where, E_{CH4} is the seasonal amount of CH₄ uptake (g CH₄ m⁻²), F_i and F_{i+1} are the CH₄ fluxes of the *i*-th and *i* + 1-th measurement (mg CH₄ m⁻² h⁻¹), the term of $(t_{i+1} - t_i)$ denotes the days between two adjacent days of the measurements, *n* is the total times of the measurements, and 24 is the number of hours per day.

Parallel to the flux measurements, soil temperature was recorded at the soil surface, 5 cm, and 10 cm belowground. Soil moisture was recorded at the 10 cm depth. Due to the proximity of the four subplots (<10 m apart) and the simultaneous collection of air samples, soil temperature in the four subplots is considered to be the same. Volumetric soil moisture was transformed to water-filled pore space (WFPS): WFPS = volumetric soil moisture / (1 – bulk density / 2.65) (C. Jiang et al., 2010).

2.4. Field sampling and measurements

On the 15th day of each month during the two growing seasons, cores of mineral soil nearby the gas chambers were taken at 10 cm intervals to a depth of 50 cm using an auger (2.5 cm in diameter). Four core sets were collected at each plot and like depths were homogenized to obtain a sample. Soils were immediately passed through a 2 mm sieve to remove roots, gravel and stones. Subsamples of each depth interval were extracted to determine $\rm NH_4^+-N$ and $\rm NO_3^--N$ concentrations using a continuous-flow autoanalyzer

(Bran Luebbe, Germany). Soil inorganic N storages (SIN, g m^{-2}) at each plot were calculated using the following equation:

$$SIN = \sum_{i=1}^{5} C_i \times BD_i \times 0.1$$

where, *SIN* is the storage of inorganic N at a plot (mg m⁻²), C_i is the concentration of inorganic N in the *i*-th horizon (mg kg⁻¹), BD_i is soil bulk density of the *i*-th horizon (g cm⁻³), and 0.1 is the conversion factor.

Soil pH was determined in a 1:2 soil:water suspension using a standard pH meter (Mettler Toledo, Switzerland). In addition, aboveground biomass at each plot was measured by clipping vegetation at the ground level, drying plants at 60 $^{\circ}$ C to a constant mass, and weighing.

2.5. Statistical analysis

Repeated measurement analysis of variance was used to test the differences of soil temperature, WFPS, pH, inorganic N storages, aboveground biomass and CH₄ fluxes among the different forms and rates of N addition. Comparisons of the means were conducted using the Tukey's HSD test. Simple and multiple stepwise regression analyses were used to examine the relationships between soil CH₄ fluxes and related environmental variables. All statistical analyses were conducted using the SPSS software package (version 16.0). Statistical significant differences were set with *p* values < 0.05 unless stated otherwise.

3. Results

3.1. Soil CH₄ uptake

The alpine meadow ecosystem behaved as a net sink of atmospheric CH₄ during the growing seasons in 2008 and 2009. Soil CH₄ fluxes varied as a single-peak pattern, and the maximum occurred in early August 2008 (Fig. 2a–c). However, in 2009, the seasonality of soil CH₄ fluxes weakened (Fig. 2a–c). In the control plots, the CH₄ uptake fluxes from the alpine meadow ecosystem averaged $31.1 \pm 3.5 \ \mu\text{g}$ CH₄ m⁻² h⁻¹, which was converted into the seasonal amount of 104.69 \pm 5.60 g CH₄ m⁻² (Table 1). N addition tended to

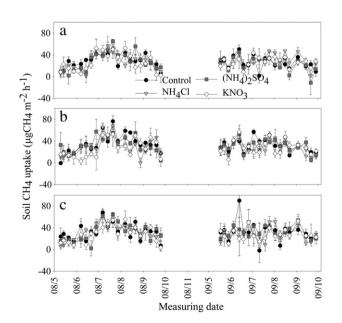


Fig. 2. Seasonal variations of soil CH_4 fluxes in low (a), medium (b), and high (c) N treatments.

inhibit soil CH₄ uptake, especially in medium N treatments (Table 1). N addition decreased the seasonal amount of CH₄ uptake by -1.9% to 18.5% without considering N addition types, with a mean of 8.6% (Table 1).

3.2. Soil temperature and WFPS

During the entire growing season, the soil surface temperature fluctuated greatly (Fig. 3a–c). Soil temperature at the 5 cm and 10 cm depths exhibited a mono-peak change, ranging from 3.2 to 15.9 °C (Fig. 3a–c). The maximum soil temperature occurred at the end of July (Fig. 3a–c). There was no significant difference in soil temperature from the surface, 5 cm or 10 cm depths among the different N addition rates (Table 1).

In contrast to soil temperature, soil WFPS exhibited an overall single-pit pattern in 2008. The high WFPS values were recorded in early May and at the end of September, and fluctuated violently in the other months (Fig. 4a–c). Among the different types of nitrogenous fertilizers, soil WFPS showed a similar trend, with low and high rates of N addition trending to decrease soil WFPS, while medium N significantly increased soil WFPS (Table 1). In addition, the effect of NO_3^- –N fertilizer on soil WFPS seemed to be stronger than those of NH_4^+ –N fertilizers (Table 1).

3.3. Soil NH_4^+ –N and NO_3^- –N pools

The pool of soil NH₄⁺–N peaked in June, and either August or September, and obviously dropped in July (Fig. 5a–c). In the control plots, soil NH₄⁺–N storage in the alpine meadow ranged from 1.83 to 7.86 g m⁻², with an average of 4.01 \pm 0.30 g m⁻² (Fig. 5a–c). N addition was seen to both deplete and accumulate soil NH₄⁺–N pool, despite the fertilizer form, but the difference between N addition plots and control plots was not significant after three-year N addition (Table 1).

The monthly variation of soil NO₃⁻–N pools was not consistent with that of soil NH₄⁺–N pools. The peaks of soil NO₃⁻–N pools occurred in June or July, and minimum values occurred in August or September (Fig. 6a–c). In the control plots, soil NO₃⁻–N pools in the alpine meadow soils ranged from 0.59 to 3.14 g m⁻², with an average of 1.47 \pm 0.12 g m⁻², which was significantly lower than that of soil NH₄⁺–N pool (Table 1). In 2008 and 2009, soil NO₃⁻–N pool accumulated in a positive trend with N addition; moreover, the cumulative effect of NO₃⁻–N fertilizer was slightly higher than those of NH₄⁺–N fertilizers (Table 1). However, the difference between N addition treatments and control was not significant (Table 1).

3.4. Aboveground biomass

Aboveground biomass was lowest in May, then gradually increased and reached its maximum in August, after which it began to decrease (Fig. 7a–c). In the control plots, aboveground biomass in the alpine meadow ranged from 147.6 to 581.3 g m⁻², with an average of 356.4 \pm 9.9 g m⁻² (Fig. 7a–c). Three levels of N addition consistently increased aboveground biomass during the growing seasons of 2008 and 2009; moreover, the positive effect of N addition on plant growth significantly increased with the N addition rate (Table 1).

3.5. Soil pH

Throughout the growing season, soil pH did not vary dramatically (Fig. 8a–c). In 2008, N addition tended to decrease soil pH, especially in the high N plots (Fig. 8a–c). In 2009, N addition significantly decreased soil pH values in the forms and levels of nitrogenous fertilizer addition (Fig. 8a–c). The decline of soil pH caused by N addition ranged from 0.07 to 0.21 units without considering N addition forms (Table 1). Moreover, the decrease of soil pH values caused by NO₃⁻–N fertilizer was higher than that caused by NH₄⁺–N fertilizers (Table 1).

The seasonal amount of soil CH ₄ uptake and relevant soil properties in different N treatments over the two growing seasons of 2008 and 2009.										
N level	N form	Seasonal CH ₄ uptake (g CH ₄ m ⁻²) ^a	Soil temperature (°C) ^a			Soil WFPS	Soil NH ₄ ⁺ -N	Soil NO ₃ ⁻ -N	Abov	
			Surface	5 cm	10 cm	$(m^3 m^{-3})^a$	pool (g m ⁻²) ^a	pool (g m ⁻²) ^a	biom (g m	
Control	Control	104.69 ± 5.60 a	$137 \pm 0.7a$	$92 \pm 0.6a$	$93 \pm 07a$	0.37 ± 0.02 h	4.01 ± 0.30 ab	147 ± 0.12 a	356.4	

N level	N form	Seasonal CH ₄ uptake (g CH ₄ m ⁻²) ^a	Soil temperature (°C) ^a			Soil WFPS	Soil NH ₄ ⁺ -N	Soil NO ₃ -N	Aboveground	Soil pH ^a
			Surface	5 cm	10 cm	$(m^3 m^{-3})^a$	pool (g m ⁻²) ^a	pool (g m ⁻²) ^a	biomass (g m ⁻²) ^a	
Control	Control	$104.69 \pm 5.60 \text{ a}$	13.7 ± 0.7 a	9.2 ± 0.6 a	9.3 ± 0.7 a	$0.37\pm0.02b$	4.01 ± 0.30 ab	1.47 ± 0.12 a	$356.4\pm9.9\mathrm{b}$	7.37 ± 0.02 a
Low N	NH ₄ Cl	106.68 ± 1.56 a	13.0 ± 0.8 a	9.2 ± 0.6 a	9.2 ± 0.6 a	$0.35\pm0.03b$	3.81 ± 0.56 b	1.53 ± 0.16 a	376.0 \pm 7.2 ab	7.29 ± 0.09 ab
Low N	(NH ₄) ₂ SO ₄	103.83 ± 6.99 a				$0.35\pm0.03c$	$4.30\pm0.61~\mathrm{ab}$	1.58 ± 0.17 a	397.1 ± 7.6 a	7.30 ± 0.07 ab
Low N	KNO ₃	104.52 ± 3.19 a				$0.34\pm0.03~{ m c}$	$3.81\pm0.56~\mathrm{b}$	1.78 ± 0.18 a	$390.6 \pm 7.3 \mathrm{~ab}$	7.24 ± 0.17 ab
Medium N	NH ₄ Cl	$85.31 \pm 9.76 \mathrm{b}$	13.5 ± 0.5 a	9.4 ± 0.6 a	9.3 ± 0.7 a	0.39 ± 0.02 ab	$4.23\pm0.60~\mathrm{ab}$	1.44 ± 0.18 a	378.3 \pm 6.7 ab	7.30 ± 0.08 ab
Medium N	$(NH_4)_2SO_4$	$93.78 \pm 6.91 \ { m ab}$				0.39 ± 0.03 ab	4.28 \pm 0.62 ab	1.65 ± 0.19 a	$398.8 \pm 6.9 \mathrm{a}$	7.32 ± 0.08 ab
Medium N	KNO3	$87.37 \pm 0.87 \text{ b}$				$0.40\pm0.02~\mathrm{a}$	$4.20\pm0.64~\mathrm{ab}$	1.68 ± 0.20 a	404.8 \pm 8.1 a	7.15 ± 0.13 b
High N	NH ₄ Cl	$94.59\pm8.22~\mathrm{ab}$	13.8 ± 0.9 a	9.6 ± 0.5 a	9.5 ± 0.7 a	0.36 ± 0.02 bc	4.54 ± 0.61 a	1.62 ± 0.18 a	$390.7\pm7.0~\mathrm{ab}$	7.28 ± 0.03 ab
High N	$(NH_4)_2SO_4$	94.17 \pm 1.18 ab				$0.35\pm0.03\mathrm{c}$	$4.15\pm0.58~\mathrm{ab}$	1.58 ± 0.17 a	$400.8 \pm 7.7a$	$7.27~\pm~0.05~ab$
High N	KNO ₃	$91.05\pm4.22~ab$				$0.34\pm0.02c$	$3.85\pm0.56~b$	1.66 ± 0.16 a	$409.8\pm7.1a$	$7.19\pm0.10~b$

^a Data are shown as means with standard errors; different lower case letters in a column indicate significant differences between treatments at *p* < 0.05.

3.6. Relationships between soil CH₄ fluxes and environmental variables

Table 1

Soil CH₄ fluxes were positively correlated with soil temperature at the 5 cm depth, while negatively correlated with soil WFPS at the 10 cm depth (Fig. 9a–b). The relationships between soil CH₄ fluxes and soil WFPS could be well fitted with an exponential decay equation (Fig. 9b). Although soil CH₄ uptakes were positively correlated with soil NO_3^- -N and NH_4^+ -N pools, only the relationships between CH_4 fluxes and soil NO_3^- -N pools were significant (Fig. 9c-d and Table 2). These results showed that soil drought and NO₃-N accumulation contributed to soil CH₄ consumption. Also, the relationships between soil CH₄ fluxes and aboveground biomass values were fitted by a quadratic equation (Fig. 9e-f), suggesting that vegetation has positive and negative effects on soil CH₄ uptake. Except for control plots, soil CH₄ fluxes at the N addition plots were positively correlated with soil pH, suggesting that soil acidification caused by N addition could inhibit soil CH₄ uptake. In the control plots, soil WFPS, temperature, aboveground biomass and NO₃⁻-N pool could explain 38.5%, 23.4%, 12.7% and 3.7% of the variation of soil CH₄ uptakes, respectively (Table 2). N addition tended to increase the contributions of soil WFPS, pH and NO_3^- pool to CH_4 uptake, which could be reflected by regression coefficient (R^2) values (Table 2). Multiple stepwise regression analysis indicated that soil CH₄ fluxes were mainly controlled by soil WFPS, followed by soil NH_4^+ -N and NO_3^- -N pools at the control plots (Table 2). However, only soil WFPS and NO₃-N pool dominated

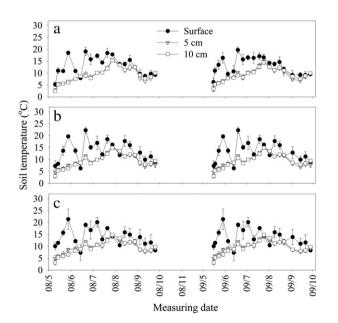


Fig. 3. Monthly variation of soil temperature in low (a), medium (b), and high (c) N treatments

soil CH₄ uptakes at the N addition treatment plots (Table 2). Therefore, our results suggest that the decreasing physical diffusion by soil WFPS rather than biochemical inhibition by other variables was responsible for the decrease in soil CH₄ uptake.

4. Discussion

4.1. Comparisons with other studies

Under natural conditions, the alpine meadow ecosystem is a sink of atmospheric CH₄. The average CH₄ uptake flux during the study period $(31.1 \pm 3.5 \ \mu g \ CH_4 \ m^{-2} \ h^{-1})$ is close to the values $(26-30 \ \mu g \ CH_4 \ m^{-2} \ h^{-1})$ reported by other studies at the same site (Cao et al., 2008; Lin et al., 2009). However, the fluxes of CH₄ uptake are lower than fluxes reported in alpine steppe grasslands in the middle Tibetan Plateau, China (63.4–70.2 μ g CH₄ m⁻² h⁻¹, Wei et al., 2012), the alpine grassland of the Tianshan Mountains. China (54.2 \pm 6.9 μ g CH₄ m⁻² h⁻¹, Li et al., 2012) and the alpine dry meadow in the Eastern Alps (41.7–87.5 μ g CH₄ m⁻² h⁻¹, Koch et al., 2007). CH_4 fluxes are significantly correlated with soil moisture (Fig. 9), and the difference in moisture availability is likely to be the most important driver of soil CH₄ fluxes in these alpine grasslands (Sjögersten et al., 2012). The soil moisture in the Haibei alpine meadow (37%) was higher than those in the above study sites (less than 30%), which is the main reason for the lower soil CH₄ uptake.

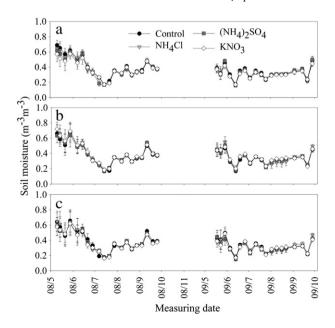


Fig. 4. Monthly variations of soil WFPS in low (a), medium (b), and high (c) N treatments.

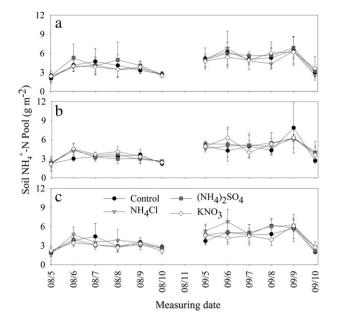


Fig. 5. Monthly variations of soil $\mathsf{NH}_4^+\mathsf{-N}$ pool in low (a), medium (b), and high (c) N treatments.

Similar to many N addition experiments in grassland ecosystems (Domingues et al., 2007; Liebig et al., 2008; Mosier et al., 1998), CH₄ uptake in the alpine meadow system is significantly inhibited by N addition over the short-term. This differs from some studies saying that N addition does not significantly affect or promote soil CH₄ uptake (Phillips and Podrebarac, 2009; Sawamoto et al., 2010; Van den Pol-van Dasselaar et al., 1999). Moreover, the rates of N addition in our study were lower than those in the above studies, suggesting that the alpine meadow ecosystem is very sensitive to increased N deposition. The fact that N addition is inhibiting the CH₄ uptake implies that the Tibetan Plateau may remove less CH₄ under future increased nitrogen deposition conditions.

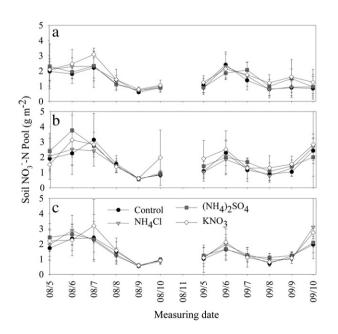


Fig. 6. Monthly variations of soil $NO_3^-\text{-}N$ pool in low (a), medium (b), and high (c) N treatments.

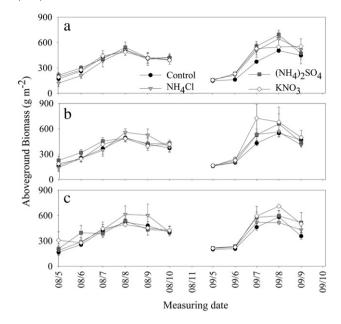


Fig. 7. Monthly variations of above ground biomass in low (a), medium (b), and high N (c) treatments.

4.2. Effects of soil WFPS on CH₄ uptake

Theoretically, the variation of soil moisture content depends on the dynamic balance between the input by precipitation and the loss by ecosystem evapotranspiration in semiarid alpine meadow ecosystems. In the study, precipitation was the same among the plots, and thus the differences in soil moisture content among the different N addition plots could be attributed to differences in ecosystem evapotranspiration among the plots. Ecosystem evapotranspiration is positively correlated with soil moisture (St Clair et al., 2009). N fertilizer application generally not only increases plant primary production and decreases species diversity, but also increases evapotranspiration (Sonnleitner et al., 2001). Our study showed that low and high N addition tended to decrease soil moisture, which could be related to the increase of ecosystem evapotranspiration. However, the responses of soil moisture

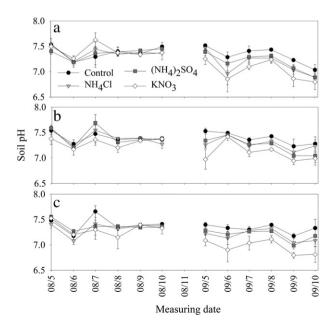


Fig. 8. Monthly variations of soil pH in low (a), medium (b), and high (c) N treatments.

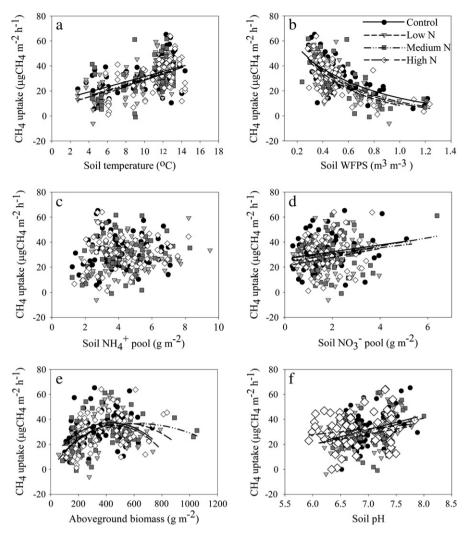


Fig. 9. Relationships between soil CH₄ fluxes and soil temperature, WFPS, inorganic N pools, pH and aboveground biomass.

to N addition are ambiguous—including positive (Fang et al., 2012), negative (Inouye, 2006), and neutral (Xia et al., 2009) results. Therefore, the driving mechanism behind this pattern needs to be examined further.

Because diffusion of CH₄ in water is 104 times slower than in air (Marrero and Mason, 1972), soil moisture controls the mass flow of air and diffusion of atmospheric CH₄ into the soil by altering the WFPS of soils (Domingues et al., 2007; Lin et al., 2009). The temporal variation in CH₄ uptake from each plot was consistent with that of soil WFPS (Figs. 2 and 4). Moreover, the fluxes of CH₄ uptake from N addition plots and control plots were significantly related to soil WFPS (Fig. 8b and Table 2). N addition usually stimulates vegetation growth in terrestrial ecosystems, and the subsequent changes in evapotranspiration enhance or lower soil WFPS, leading to higher or lower diffusion of CH₄ and O₂ into the soil (Veldkamp et al., 2001). In our study, medium N addition significantly increases soil WFPS, which corresponds to the significant decrease in soil CH₄ uptake (Table 1). Compared with other variables, soil WFPS plays a more important role in CH₄ uptake based on multiple regression results. This implies that physical diffusion dominates the direction and magnitude of the CH₄ uptake variation caused by N addition. Moreover, the determination coefficient (R^2) between CH₄ uptake fluxes and soil WFPS values increases from 38.5% at control plots to 43.4-47.2% at N addition plots (Table 2), suggesting an increasing contribution of soil WFPS to the variation of CH₄ uptake induced by N addition.

4.3. Effects of soil NH_4^+ –N variation on CH_4 uptake

Previous studies have shown that CH₄ oxidation in grassland soils can be inhibited by the addition of NH₄⁺-N fertilizers (Chan and Parkin, 2001; Dittert et al., 2005; Jacinthe and Lal, 2006), probably because (1) NH_4^+ competes with CH_4 for the same active site on the CH₄ monooxygenase of methanotrophs (Carlsen et al., 1991). (2) NH_4^+ oxidizes to the intermediate hydroxylamine (NH_2OH) by CH_4 monooxygenase, or to its further oxidation by other enzyme systems of the methanotrophs to the end product nitrite (NO_2^-) . Hydroxylamine and nitrite in turn might lead to intoxication of CH₄ oxidizers (Schnell and King, 1994). (3) Indirect effects of N-treatments, such as high osmotic pressure, could have killed methanotrophs (Bodelier and Laanbroek, 2004). Generally, the oxidation of methanotrophic bacteria is optimally active at low osmotic stress (Saari et al., 2004). (4) N addition probably increases soil CH₄ emission to offset CH₄ uptake. Higher litter input under N enrichment alleviates C limitation to microbes (Bodelier and Laanbroek, 2004). As a result, the activities of methanogenic archaea are enhanced and more CH₄ is produced (Bodelier and Laanbroek, 2004).

However, in this study, it seems that decreased CH_4 uptake is likely to be unrelated to any competitive inhibition of monooxygenase by NH_4^+ and toxic inhibition by hydroxylamine or nitrite. The relatively insignificant or positive effect of NH_4^+ on CH_4 uptake observed here may be attributed to the following aspects. First, the alpine meadow

Table 2
Regression model between soil CH ₄ fluxes and environmental variables.

N addition rates ^a	a	b	с	d	р	R^2			
$(a) F_{CH4} = a * T_s + b$									
Control	2.04	11.03			< 0.0001	0.234			
Low N	2.27	6.20			< 0.0001	0.266			
Medium N	2.26	7.04			< 0.0001	0.210			
High N	2.54	3.12			< 0.0001	0.253			
(b) <i>F</i> _{CH4} = <i>a</i> * <i>exp</i>	(−b ∗ WFPS)								
Control	72.27	1.62			< 0.0001	0.385			
Low N	86.51	2.33			< 0.0001	0.434			
Medium N	82.74	2.00			< 0.0001	0.472			
High N	85.36	2.15			< 0.0001	0.449			
(c) $F_{CH4} = a * NH2$	t + h								
Control	1 1 0				0.4341	0.0006			
Low N					0.1511	0.0154			
Medium N					0.1436	0.0165			
High N					0.3034	0.00105			
					0.5054	0.0011			
$(d) F_{CH4} = a * NO$									
Control	2.71	26.92			0.039	0.037			
Low N	3.54	20.77			0.029	0.042			
Medium N	3.76	22.63			0.010	0.062			
High N	2.20	26.77			0.014	0.057			
(e) $F_{CH4} = a * AB^2$	+b * AB + c	:							
Control	-0.0001	0.128	9.71		0.003	0.127			
Low N	-0.0001	0.104	7.22		0.008	0.085			
Medium N	-0.0001	0.073	13.31		0.005	0.118			
High N	-0.0001	0.128	2.851		0.009	0.161			
(f) $F_{CH4} = a * pH$		0.250			0.100	0.020			
Control	4.67	-0.258			0.109	0.020			
Low N	8.19	-29.08			0.011	0.066			
Medium N	10.54	-43.32			0.004	0.087			
High N	11.38	-47.83			0.002	0.103			
(g) $F_{CH4} = a * WFPS + b * NH_4^+ + c * NO_3^- + d$									
Control	-44.39	-1.72	2.13	58.48	< 0.001	0.433			
Low N	-53.22		3.97	47.45	< 0.001	0.458			
Medium N	-44.61		2.03	49.44	< 0.001	0.430			
High N	-46.05		2.62	49.43	< 0.001	0.428			

 $^a~T_s$ is soil temperature at 5 cm depth, WFPS is soil water-filled pore space at 10 cm depth, NH_4^+ and NO_3^- are soil NH_4^+ –N and NO_3^- –N storages at 50 cm depth, and AB is aboveground biomass.

soil in our study site is N-limited and has a strong capacity to immobilize exogenous NH_4^+ (Song et al., 2007). The NH_4^+ immobilization by soil organic and mineral materials rapidly removes applied NH_4^+ , which would protect methanotrophs from exposure to NH_4^+ and accordingly mitigate the inhibition of methanotrophs. Second, the increasing nitrification by N addition would to some degree alleviate the inhibition of inorganic N to CH_4 oxidation (Chan et al., 2005). Although NH_4^+ -oxidizing bacteria are capable of oxidizing CH₄ at rates lower than methanotrophs, CH₄ oxidation is dominated by nitrifiers (Steudler et al., 1996). Thus, N addition enhances the availability of NH₄⁺ to nitrifiers, which would accordingly decrease the extent to which CH₄ consumers are exposed to NH_4^+ (Chan et al., 2005). Finally, the ratio of $NH_3:NH_4^+$ in soils is decreased by N addition. Gulledge et al. (1997) suggested that NH₃, rather than NH⁺, was the key determinant of this inhibition, because the decreased soil pH induced by N addition would decrease the ratio of NH₃:NH₄⁺ of soils. This is likely to decrease the degree of inhibition linked to N addition; however, no such clear regulation of NH₄⁺ inhibition through pH has so far been identified (Gulledge et al., 1997; Hütsch, 1996).

4.4. Effects of soil NO_3^- –N variations on CH_4 uptake

Similar with NH_4^+ , NO_3^- also showed a very strong inhibitory effect in some forest ecosystems (Rigler and Zechmeister-Boltenstern, 1999). Moreover, some studies suggest that NO_3^- may have a greater importance in the inhibition of CH_4 oxidation in forest soils than that attributed to NH_4^+ (Wang and Ineson, 2003; Xu and Inubushi, 2004; Xu and Inubushi, 2007). Generally, NO_3^- has been found inhibitory only in very high concentrations, which likely give rise to osmotic effects (Bodelier and Laanbroek, 2004). The other reason is that added NO_3^- and NO_2^- produced via NO_3^- reduction in anaerobic 'microsites' are probably toxic to CH_4 -oxidizing bacteria (Schnell and King, 1994; Xu and Inubushi, 2004; Xu and Inubushi, 2007).

However, our study showed that NO₃⁻ accumulation could significantly promote CH₄ uptake in soil (Fig. 9 and Table 2). Bodelier and Laanbroek (2004) proposed a schematic explanation about the stimulation of CH_4 oxidation by NH_4^+ or NO_3^- addition to soils. For one thing, atmospheric methanotrophic bacteria are limited by N because they have a relatively high N requirement during C assimilation (Megmw and Knowles, 1987). N addition enhances soil mineralization rates and C and N availability for soil microbes (Rigler and Zechmeister-Boltenstern, 1999). Besides utilizing CH₄ in the atmosphere and in soil pores, it has been demonstrated that atmospheric CH₄ consumers can profit from and even depend on some non-methane substrates such as methanol, formate, and acetate (Bodelier and Laanbroek, 2004; Jensen et al., 1998). The improved availability of these C sources would promote CH₄ consumption by aerobic soils. Also, during the rainy season, the alpine meadow soils may become partially anoxic following high precipitation events and begin producing CH₄. The methanotrophic bacteria can profit from this enhanced CH₄ flux and grow when sufficient N is present (Bodelier and Laanbroek, 2004). However, in our study, N addition decreased overall soil CH₄ uptake. The promotion produced by NO₃⁻ accumulation is almost masked by the inhibition associated with decreasing WFPS.

4.5. Effects of other associated ions on CH₄ uptake

Besides NH_4^+ or NO_3^- , the associated ions of nitrogenous salts can also partly explain the different inhibition patterns. Some studies suggest that salt-effect may be a more reasonable explanation for reduced CH_4 oxidation than the specific NH_4^+ or NO_3^- effect (Borken and Brumme, 2009; King and Schnell, 1998; Whalen, 2000). A laboratory study showed that high concentration of Cl⁻ has a strong inhibitory effect on CH₄ oxidation by comparing the difference in ability to do so between N-salts (NH₄Cl and NaNO₃) and non-N-salt (NaCl) (Whalen, 2000). In the field, however, various N treatments $((NH_4)_2SO_4)$, NH₄-acetate, urea, NH₄Cl, NaNO₃, NH₄NO₃ and KNO₃) added at a same rate of 30 kg N ha⁻¹ had little or no effect on CH₄ uptake within one year (Borken and Brumme, 2009). The rates of N addition (10-40 kg N ha⁻¹ yr⁻¹) in our study are comparable to that of Borken and Brumme (2009), so added K^+ , SO_4^{2-} and Cl^- could have little contribution to the decrease in CH₄ uptake. Also, the K⁺ can exchange NH_4^+ from the exchange sites in the soil and thus released NH_4^+ could then inhibit CH₄ oxidation. If K⁺ was the responsible component of the added salt in the current study, ion exchange should result in a higher NH₄⁺ concentration. However, the soil NH₄⁺ pool did not accumulate significantly, and even declined in the low and high KNO₃ addition plots (Table 1). Thus, the salt effects could not be a main reason for the CH₄ uptake in our study.

4.6. Effects of vegetation on CH₄ uptake

Plant communities affect soil CH_4 uptake through the following three ways: (1) increased N input to grasslands can increase soil moisture among the fine root biomass (Magill et al., 1997), which may decrease air-filled porosity and limit rates of diffusion and atmospheric CH_4 oxidation. Our data on soil WFPS and aboveground biomass partly supports this deduction (Figs. 4 and 7). (2) Plant-derived CH_4 emission can indirectly offset the CH_4 uptake of the entire ecosystem (Cao et al., 2008; Ferretti et al., 2007; Keppler et al., 2006; Keppler et al., 2008). However, serious debates and questions on CH_4 emission by living plants under aerobic conditions are raised. Dueck et al. (2007) reported that there was no evidence for substantial aerobic CH_4 emission by plants. Moreover, aerobic CH_4 emission by plants is highly uncertain, and depends on species (Cao et al., 2008; Kirschbaum and Walcroft, 2008; Wang et al., 2007). Based on regression analysis results, CH_4 emission by alpine vegetation could have a small effect on ecosystem net CH_4 flux, although a significant relationship between CH_4 uptake and aboveground biomass is observed (Table 2 and Fig. 9e).

4.7. Soil acidification and CH₄ uptake

N addition can, to some degree, result in soil acidification in the alpine meadow, and the NO₃-N fertilizer seemed to have a stronger effect than the NH_4^+ –N fertilizer. Generally, soil H⁺ mainly originates from acid deposition, microbial decomposition of organic matter, and soil nitrification (Arnold et al., 1994; Sun et al., 2006). The oxidation of soil NH₄⁺ to NO₃⁻ (NH₄⁺ + 2O₂ \rightarrow 2H⁺ + NO₃⁻ + H₂O) can produce H⁺, so ammonium fertilizer application will directly result in soil acidification. Soil acidification caused by $NO_3^- - N$ fertilizer addition is attributed to the exchange between the accompanied cations, such as K⁺ and H⁺, on soil particle exchange sites (Fang et al., 2012; Gao et al., 2013). Compared with NH₄Cl and (NH₄)₂SO₄, the effects of KNO₃ fertilizer on soil acidification were stronger, indicating the different effects of NH_4^+ and NO_3^- on soil acidification. The added NH_{4}^{+} is partly absorbed by plants, immobilized by soil microbes, fixed by clay minerals and lost through ammonia volatilization. Only the NH₄⁺ involved in the N mineralization and nitrification can produce H^+ . On the contrary, most of the added NO_3^- can be replaced by an H⁺ due to negligible leaching and denitrification.

In some forest soils, negative effects of soil acidification on soil physical parameters and bioturbation have the potential to reduce CH_4 uptake (Borken and Brumme, 2009). However, stepwise regression analysis showed that pH was not a key factor in the control of CH_4 uptake, suggesting that: (1) soil pH plays a minor role in determining CH_4 oxidation compared with other variables, and (2) methanotrophs in the alpine meadow soils can tolerate low pH (Saari et al., 2004).

5. Conclusions

This study emphasizes the appropriateness of using realistic levels rather than saturating levels of N-inputs in attempts to simulate increased N deposition. Our study suggested that low rates of N addition had significantly reduced the CH₄ sink of alpine meadow on the Qinghai-Tibetan Plateau over the short term. This indicates that the alpine meadow has low potential to consume more CH₄ under future elevated N deposition conditions on the Tibetan Plateau. Soil CH₄ uptake was primarily driven by soil moisture. The observed reduction in CH₄ uptake induced by N addition may mainly result from the decrease in physical diffusion, which is different from those in N-rich ecosystems or in high-dose N addition experiments. Overall, N is a regulatory factor of CH₄ uptake in the alpine meadow ecosystems. To elucidate the complex mechanisms by which inorganic N affects soil CH₄ uptake, further investigation should be carried out to determine the biochemical inhibition effects of NH₄⁺ and NO₃⁻ accumulation on soil methanotrophic bacteria.

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